

In re Appln. of Waldmann et al.  
U.S. Patent Application No. 08/478,748



*SPECIFICATION AMENDMENTS*

Please amend the specification as indicated on the marked-up versions of pages 7, 8, and 9 which follow.

diminished 11-Kb EcoRI band as well as one nongermline and ( $\rightarrow$ ) that identified a monoclonal pattern of  $Tcr\beta$  gene arrangement. Furthermore, there was a diminution of the 8.0-Kb *Hind*III digest that reflects a monoclonal  $Tcr\beta$  pattern of gene rearrangement as well. This Southern blot pattern indicates that one  $Tcr\beta$  allele in the leukemic clone rearranged to  $C\beta 1$ , whereas the other allele rearranged to  $C\beta 2$ . Digests of patient DNA obtained in remission following  $^{90}Y$  anti-Tac therapy did not reveal the two non-germline bands, thus confirming the elimination of the circulating monoclonal leukemic cell population. In the schematic diagram of the germline arrangement of the  $Tcr\beta$  gene, we indicate the locations of the EcoRI (E) and *Hind*III (H) restriction endonuclease sites as well as the  $C\beta$  regions recognized by the cDNA probe used.

Fig. 5. Southern blot analysis of HTLV-I proviral integration in *Pst*I and EcoRI digests of DNA obtained from the peripheral blood mononuclear cells of Patient 7. (A) There are no EcoRI restriction sites within the HTLV-I genome. Therefore the generation of a band identifying a restriction length fragment containing HTLV-I depends on the recognition of EcoRI sites in host DNA adjacent to viral integration. Clonal integration of the complete virus is indicated by a band in an EcoRI digest that is larger than 9 Kb, the size of viral genome. The presence of a single band ( $\rightarrow$ ) demonstrable in the EcoRI digest of mononuclear cell DNA for Patient 7 during active disease demonstrates monoclonal viral integration into the leukemic cell DNA of the patient. This band was no longer evident on EcoRI digests obtained during  $^{90}Y$  anti-Tac induced complete remission. (B) There are multiple *Pst*I restriction sites within the complete HTLV-1 genome that on digestion of both polyclonally and monoclonally integrated HTLV-1 yield three bands of 1.2, 1.8, and 2.5 Kb. In digests of Patient 7 mononuclear cell DNA obtained

during active disease there are two additional bands (→) indicating monoclonal viral integration that identify one *Pst*I site within HTLV-1 and another in host DNA adjacent to the virus. This pattern is the hallmark of the presence of clonally integrated HTLV-1 in the leukemic cell DNA. No bands reflecting residual HTLV-1 were demonstrable in the *Pst*I digest of the DNA of circulating cells obtained from the patient on day 818 following initiation of therapy, supporting the view that this patient was in a complete remission. Each lane contains 10 µg of genomic DNA. Ethidium bromide staining confirmed that equivalent quantities of genomic DNA were present in each lane. A schematic diagram of the virus indicating *Eco*RI and *Pst*I restriction endonuclease sites is shown below.

Fig. 6-5. CAT scan of thorax of Patient 4 before treatment (top) and after two cycles of <sup>90</sup>Y anti-Tac therapy (bottom). There was a marked reduction in the size of the axillary lymph nodes in the scan obtained during the period when the patient was in an <sup>90</sup>Y anti-Tac therapy-induced partial remission.

Fig. 7-6. <sup>111</sup>In anti-Tac imaging studies of Patient 1 prior to treatment and at the time of the fourth treatment with <sup>90</sup>Y anti-Tac when the patient was in a complete remission. Prior to therapy <sup>111</sup>In anti-Tac was deposited in sites of malignant T-cell infiltration of the skin of the hands, whereas no such deposition was evident at the time of the fourth study confirming the complete remission.

Fig. 8. <sup>111</sup>In anti-Tac anterior whole body scans from Patient 4 obtained at 48 hours post-tracer administration. Left panel shows accumulation in involved axillary, cervical, inguinal, and hilar nodes in the images obtained at the time of the initial therapeutic infusion. The patient received 5 mCi of <sup>111</sup>In and 10 mCi

of  $^{90}\text{Y}$  anti-Tac with a total of 10 mg of antibody. The right panel was obtained 6 weeks after the patient's first therapy. The imaging dose was identical to the first dose. The  $^{111}\text{In}$  anti-Tac study revealed a marked decrease in tumor size and more prolonged circulation of the tracer-labeled antibody, which was associated with the decreased tumor burden.

Fig. 9-7.(A) Effect of  $^{90}\text{Y}$  anti-Tac therapy on the absolute number of Tac-expressing ATL leukemic and normal T cells/mm<sup>3</sup> of Patient 7.  $^{90}\text{Y}$  anti-Tac monoclonal antibody was administered i.v. to the patient at the doses and on the days indicated by the arrows (→). The patient initially had 27,875 circulating Tac-expressing malignant cells/mm<sup>3</sup>(--) The patient received 50 mCi of  $^{90}\text{Y}$  anti-Tac during the first 410 days of therapy in divided doses. By day 300 following initiation of therapy, the patient had undergone a complete remission that has been maintained for the over 800-day period of observation. There was an initial modest reduction in the number of normal T cells (○-) (normal T cells are CD7<sup>+</sup>CD25<sup>-</sup>). However, the number of these normal T cells subsequently returned to pretreatment levels during the remaining period when the patient was in a sustained complete remission. (B) Effect of  $^{90}\text{Y}$  anti-Tac therapy on the serum concentration of sIL-2R $\alpha$  of the same patient. The serum sIL-2R $\alpha$  level of the patient prior to therapy was 2,938 units/ml. The concentration sIL-2R $\alpha$  returned to normal or below normal levels following therapy confirming the complete remission.

Fig. 10-8. A Kaplan-Meier plot (1958 *J. Am. Stat. Assoc.* 53:457) of event-free survival (surviving patients without progressive disease) comparing patients treated with unmodified anti-Tac (---) with those receiving  $^{90}\text{Y}$  anti-Tac (-).